## AAV Sequence Analysis

Sample:  $m84028\_231102\_045756\_s2$ 

Date: 2024-08-16

## ${\bf Contents}$

Read type classification	2
Definitions	2
Assigned types by read alignment characteristics Single-stranded vs self-complementary frequency	5
Distribution of read lengths by assigned AAV types	6
Assigned AAV read types detailed analysis Assigned AAV types (top 20)	
Flip/flop considerations Flip/flop configurations, scAAV only	7
Distribution of read length by subtype	8
AAV mapping to reference sequence Gene therapy construct	ę G
Distribution of non-matches by reference position	10
Methods Citations	11 12

## Read type classification

#### Definitions

Assigned Type	Definition
$\operatorname{scAAV}$	Self-complementary AAV where one half of the payload region is a reverse complement of the other, resulting in an intra-molecular double-stranded DNA template. A sequencing read is inferred as scAAV if it has both a primary and supplementary alignment to the vector genome.
ssAAV	AAV where the resulting DNA template is expected to be single-stranded, as opposed to self-complementary. A sequencing read is inferred as ssAAV if it has a single alignment to the AAV genome and no complementary secondary alignment.
other	Read consists of a fragment mapping to the vector but with unexpected polarities (e.g. +,-,- or +,+,+) and cannot be well-defined at the moment. This means that the algorithm was not able to distinguish the read either as ssAAV or scAAV by the definitions above. Usually this means there are multiple supplementary alignments on the vector region and/or it's a weird molecule.
host	Read originates from the host genome that is given (e.g. hg38, CHM13).
repcap	Read originates from the repcap plasmid. The Rep gene encodes four proteins (Rep78, Rep68, Rep52, and Rep40), which are required for viral genome replication and packaging, while Cap expression gives rise to the viral capsid proteins (VP; VP1/VP2/VP3), which form the outer capsid shell that protects the viral genome, as well as being actively involved in cell binding and internalization.
helper	Read originates from the helper plasmid. In addition to Rep and Cap, AAV requires a helper plasmid containing genes from adenovirus. These genes (E4, E2a and VA) mediate AAV replication.
chimeric	Read consists of fragments that map to one or more "genomes" (e.g. vector and host; helper and repcap).

Note: Even though ssAAV distinguishes one ITR as the wildtype (wtITR) and the other as the mutated ITR (mITR), we will still refer to them as "left ITR" and "right ITR". For example, "left-partial" would be equivalent to "mITR-partial" in the case where the mITR is the left ITR based on the given genomic coordinates.

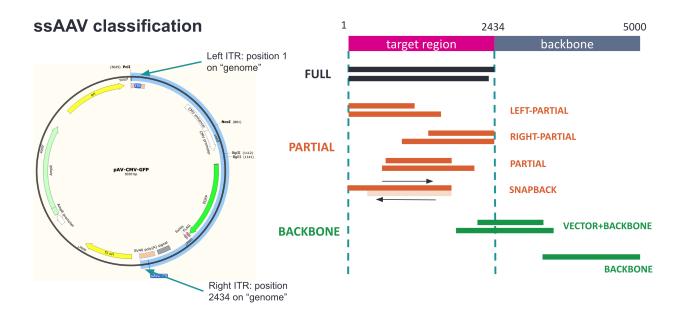


Figure 1: Single-stranded AAV types (ssAAV). ssAAV reads must fully map within the vector genome (which can contain the backbone, beyond the ITR region) consisting of a single (primary) alignment. "ssAAV-full" reads must cover from left ITR to right ITR. "ssAAV-left-partial" contain the left ITR but are missing the right ITR. "scAAV-vector+backbone" map partially within the ITR and partially to the backbone.

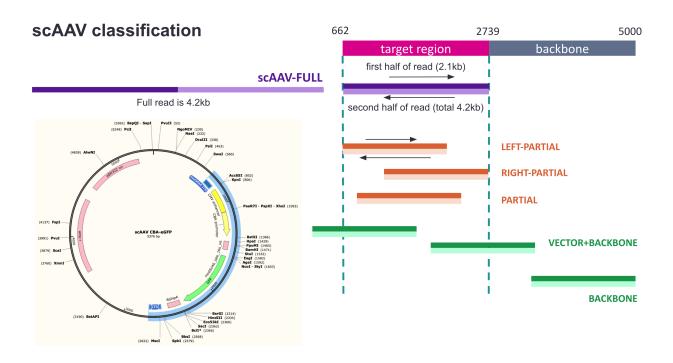
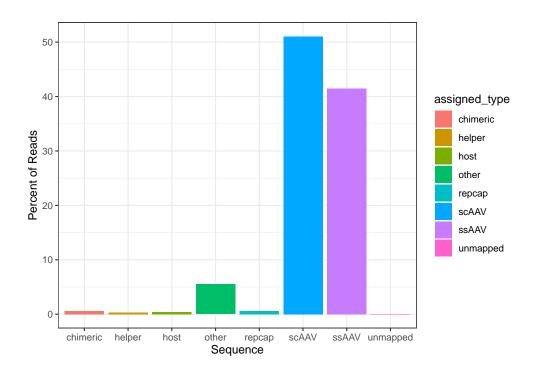
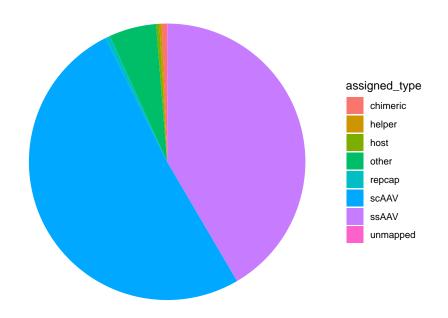


Figure 2: Self-complementary AAV types (scAAV). scAAV reads must fully map within the vector genome (which can contain the backbone, beyond the ITR region) consisting of both primary and supplementary alignments. The definitions of full, partial, and backbone reads are the same as in ssAAV.

## Assigned types by read alignment characteristics

Assigned Type	Count	Frequency (%)
scAAV	1,387,444	51.07
ssAAV	1,127,606	41.50
other	149,756	5.51
chimeric	16,879	0.62
repcap	15,697	0.58
host	10,477	0.39
helper	7,483	0.28
unmapped	1,512	0.06

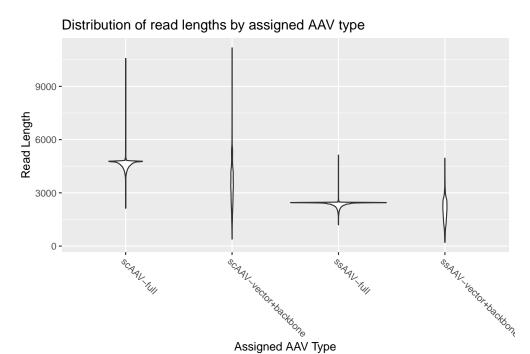




Single-stranded vs self-complementary frequency

Assigned Type	Count	Frequency in AAV (%)	Total Frequency (%)
scAAV	1,387,444	55.17	51.07
ssAAV	1,127,606	44.83	41.50

## Distribution of read lengths by assigned AAV types



## Assigned AAV read types detailed analysis

Assigned AAV types (top 20)

Assigned Subtype	Count	Freq. in AAV (%)	Total Freq. (%)
full	553,634	22.01	20.38
left-partial	188,741	7.50	6.95
right-partial	316,265	12.57	11.64
partial	64,645	2.57	2.38
backbone	1,274	0.05	0.05
vector+backbone	3,047	0.12	0.11
full	370,134	14.72	13.62
left-partial	158,135	6.29	5.82
right-partial	431,999	17.18	15.90
partial	101,804	4.05	3.75
backbone	20,950	0.83	0.77
vector+backbone	14,070	0.56	0.52
full right-partial	70,469	2.80	2.59
right-partial partial	57,682	2.29	2.12
full left-partial	44,441	1.77	1.64
	full left-partial right-partial partial backbone vector+backbone full left-partial right-partial partial backbone vector+backbone full left-partial right-partial partial backbone	full       553,634         left-partial       188,741         right-partial       316,265         partial       64,645         backbone       1,274         vector+backbone       3,047         full       370,134         left-partial       158,135         right-partial       431,999         partial       101,804         backbone       20,950         vector+backbone       14,070         full right-partial       70,469         right-partial partial       57,682	full       553,634       22.01         left-partial       188,741       7.50         right-partial       316,265       12.57         partial       64,645       2.57         backbone       1,274       0.05         vector+backbone       3,047       0.12         full       370,134       14.72         left-partial       158,135       6.29         right-partial       431,999       17.18         partial       101,804       4.05         backbone       20,950       0.83         vector+backbone       14,070       0.56         full right-partial       70,469       2.80         right-partial partial       57,682       2.29

Assigned Type	Assigned Subtype	Count	Freq. in AAV (%)	Total Freq. (%)
scAAV	left-partial partial	24,829	0.99	0.91
scAAV	partial right-partial	20,383	0.81	0.75
scAAV	left-partial full	16,566	0.66	0.61
scAAV	right-partial full	15,334	0.61	0.56
scAAV	left-partial right-partial	12,259	0.49	0.45

#### Definitions

Assigned Subtype	Definition
full	Read alignment includes the entire ITR-to-ITR target vector sequence.
left-partial	Read aligns to a fragment of the vector originating from the left (upstream) ITR of
	the vector while not covering the right ITR.
right-partial	Read aligns to a fragment of the vector originating from the right (downstream) ITR
	of the vector while not covering the left ITR.
partial	Read aligns to a fragment of the vector originating from within the ITR sequences.
vector+backbone	Read aligns to a fragment including the vector as well as plasmid backbone sequence.
backbone	Read aligns to a fragment originating solely from the plasmid backbone sequence.
snapback	Read consists of a double-stranded, sub-genomic fragment including only one ITR
	and read alignments in both (+) and (-) polarities. (ssAAV only)

## Flip/flop considerations

Term	Definition
Flip/Flop	One ITR is formed by two palindromic arms, called B–B' and C–C', embedded in a larger one, A–A'. The order of these palindromic sequences defines the flip or flop orientation of the ITR. (Read more)

No flip/flop analysis results available to display.

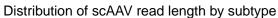
#### Flip/flop configurations, scAAV only

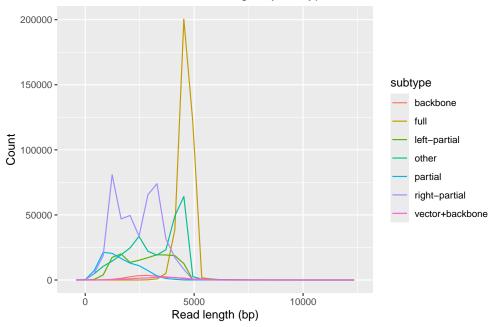
No scAAV flip/flop analysis results available to display.

### Flip/flop configurations, ssAAV only

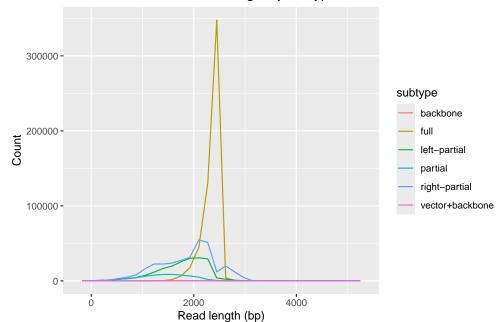
No ssAAV flip/flop analysis results available to display.

## Distribution of read length by subtype



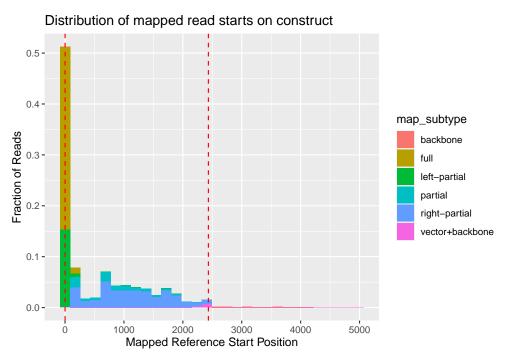


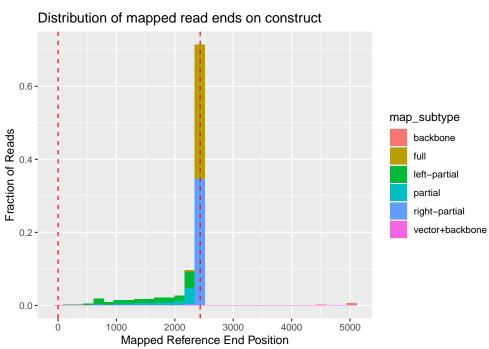
#### Distribution of ssAAV read length by subtype

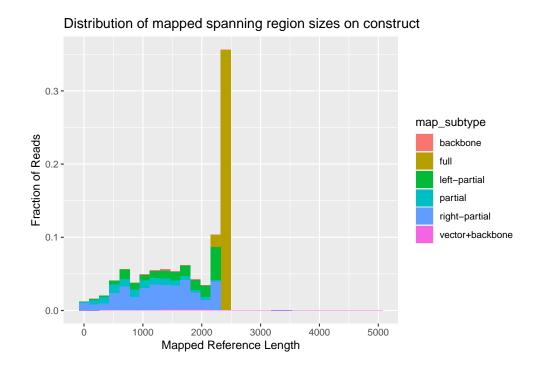


## AAV mapping to reference sequence

#### Gene therapy construct

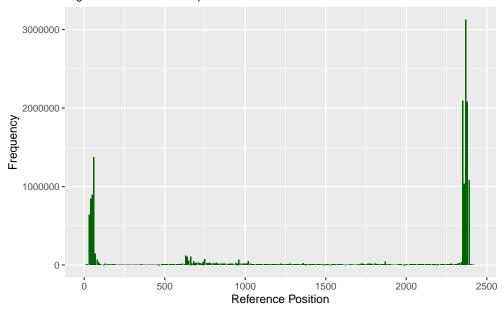




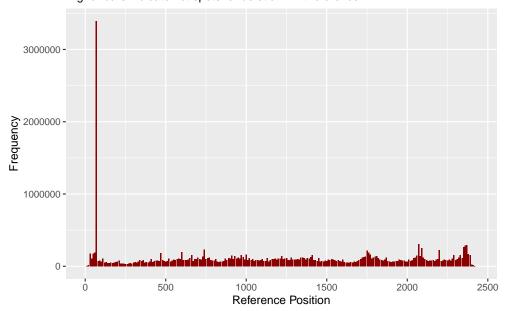


## Distribution of non-matches by reference position

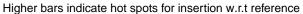
# Substitutions by reference position Higher bars indicate hot spots for substitutions w.r.t reference

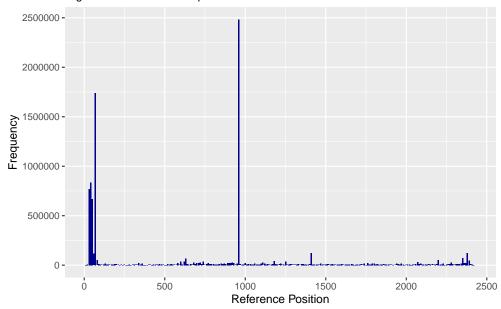


## Deletions by reference position Higher bars indicate hot spots for deletion w.r.t reference



#### Insertions by reference position





#### Methods

This report was generated by an automated analysis of long-read sequencing data from adeno-associated virus (AAV) products. The sequencing data should be from the PacBio sequencer run in AAV mode, or equivalent circular consensus sequencing (CCS) reads (Travers et al., 2010). Reads are aligned to the AAV, packaging, and masked host reference sequences using Minimap2 (Li, 2018).

In this analysis, aligned sequencing reads were filtered for quality to include primary alignments and reads with mapping quality scores greater than 10. The alignment coordinates and orientation of reads passing

these filters were then compared to the annotated vector region in the reference sequence, which comprises the left and right ITRs and the genomic region between them, to assign each read to a type and (for AAV reads) subtype classification according to the definitions above.

#### Citations

- 1. Travers, K. J., Chin, C.-S., Rank, D. R., Eid, J. S. & Turner, S. W. A flexible and efficient template format for circular consensus sequencing and SNP detection. Nucleic Acids Research 38, e159–e159 (2010).
- 2. Li, H. Minimap2: Pairwise alignment for nucleotide sequences. Bioinformatics 34, 3094–3100 (2018).